Standard Operating Procedures
Lentivirus and Lentivirus Vector Usage in Rodents

The procedures described below should be utilized only when the common, well described replication incompetent retroviral vectors (and packaging systems) are inoculated into rodents.

Inoculation:

- Initial delivery of the lentivirus vector must be performed within a biological safety cabinet under biosafety level 2 (BL2) conditions.

- Animal must be anesthetized or restrained in a manner that prevents sudden movements and potential auto-inoculation.

- Luer-lok syringes must be used.

- The site of inoculation should be cleaned thoroughly with 70% EtOH before placing the animal in a clean disposable cage.

- Work surfaces should be cleaned with 10-20% bleach.

Housing:

- After injection, place the animal in a clean disposable cage for a recovery period of 8-20 hours. Recovery is conducted in the BL2 facility.

- Label the cage with a biohazard label, virus utilized, route of injection, date of injection, and personnel contact information.

- After 8-20 hours, the animal is transferred to a FBRL micro-isolator and returned to the animal facility. At this point, BL1 practices are acceptable for animal containment and handling.

- Cage, bedding, food, and water bottles utilized in the BL2 facility are placed in a bio-hazard bag and autoclaved. The biohazard bag must be removed from the autoclave by the lab personnel in a timely manner and disposed of per Brandeis Biohazardous Waste Regulations.

Carcass Disposal

- Rodent carcasses that have been injected with lentivirus should be placed in the biohazard bag in the freezer at Foster Labs. Only animals can be placed in the biohazard bag-no paper towels, gloves, etc.
Training

- All personnel working with viruses must have completed Brandeis University’s Biosafety Training.