

**Isaac J. Krauss, Ph. D.**  
Curriculum Vitae

Department of Chemistry  
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## **BIOGRAPHICAL INFORMATION**

Born and raised in Fairbanks, Alaska

## **DEGREES**

Ph. D. 2003 (Chemistry, Columbia University, Advisor James Leighton)  
M. Phil. 2002 (Chemistry, Columbia University, Advisor James Leighton)  
M. A. 2000 (Chemistry, Columbia University, Advisor James Leighton)  
B. A. 1998 (Chemistry, Stanford University, Advisor Barry M. Trost)

## **PROFESSIONAL EXPERIENCE**

**Brandeis University**, Waltham, MA (July 2008–present)

Assistant Professor of Chemistry, 2008–2015

Associate Professor, 2015–2020

Professor, 2020–present

**Roche Bioscience**, Palo Alto, CA (Summer 1996)

Summer intern in natural product synthesis

## **EDUCATION**

**Memorial Sloan–Kettering Cancer Center**, New York, NY (January 2004 – June 2008)

NIH Postdoctoral Fellow working on carbohydrate vaccines and natural product synthesis

Advisor: Professor Samuel J. Danishefsky

**Columbia University**, New York, NY (August 1998 – September 2003)

Graduate work on diastereo- and enantioselective transition-metal-catalyzed reactions

Advisor: Professor James L. Leighton

**Stanford University**, Palo Alto, CA (September 1993 – June 1998)

Undergraduate research on pi-allyl substitutions

Advisor: Professor Barry M. Trost (September 1996 – June 1998)

Teaching assistant for 4 courses in organic and analytical chemistry, including one quarter as Head TA.

**West Valley High School**, Fairbanks, AK (September 1989–June 1993)

Valedictorian

## **INDEPENDENT RESEARCH**

**Project A: Chemical Biology** My laboratory is involved in directed evolution of modified DNAs and peptides. In the DNA field, we have developed a system that allows selections of DNA aptamers decorated with large carbohydrates. We have applied this system to the directed evolution of DNA-scaffolded glycoclusters which bind to broadly-neutralizing anti-HIV antibody 2G12 with low nanomolar affinity. In a

related project, we have modified the peptide directed evolution technique, mRNA display, to enable directed evolution of glycopeptides. We have used this technique to discover glycopeptides that are very close mimics of HIV protein gp120, being recognized by antibody 2G12 with sub-nanomolar affinities. Both the glycoDNA and glycopeptide constructs are being investigated for their applications as HIV vaccines.

**Project B: Organic Synthesis** We are also interested in the development of synthetic methods and organic synthesis. Recently we have developed a method for homoallylation and homocrotylation of aldehydes using cyclopropylcarbinylboronates (cyclopropanated allyl- and crotylboronates), allowing access to 1,3-syn or anti-substituted bishomoallylic alcohols, which are difficult to access by other methods. These compounds are important for the synthesis of numerous tetrahydrofuran and tetrahydropyran-containing natural products.

### **RESEARCH EXPERIENCE (Postdoctoral and Predoctoral)**

**Postdoc in Synthesis and Chemical Biology**, Memorial Sloan-Kettering Cancer Center, New York, NY (January 2004 – June 2008)

In the laboratory of Samuel Danishefsky, developed a total synthesis of the unstable macrolide, isomigrastatin, a natural product which inhibits the migration of cancer cells and is part of a family of natural products with anti-metastatic properties.

Together with other post docs in the Danishefsky, Moore, and Massague groups at Sloan-Kettering, developed migrastatin analogs with improved anti-metastatic properties in *in vivo* metastasis models in mice.

Designed and synthesized multivalent cyclic glycopeptide mimics of an epitope on HIV glycoprotein gp120. In collaboration with Merck, these compounds were used to raise anti-carbohydrate antibodies in guinea pigs and rhesus macaques.

**Ph. D. work in Transition-Metal-Catalyzed Reactions**, Columbia University, New York, NY (January 1999 – September 2003)

In the laboratory of James Leighton, used a directing group strategy to develop branched selective rhodium-catalyzed hydroformylation, allowing a non-aldol route to anti-polypropionate motifs.

Developed easily-synthesized sulfonamide-phosphine hemi-labile ligands which induced unprecedentedly high ee's in asymmetric conjugate addition of organozinc reagents and proceeded under practical room-temperature conditions.

**Undergraduate Research in Transition-Metal-Catalyzed Reactions**, Stanford University, Palo, Alto, CA (September 1996 – June 1998)

In the laboratory of Barry Trost, Developed conditions for asymmetric alkylation of malonate and imide (C- and N-) nucleophiles with racemic gamma-hydroxybutenolide carbonates, using Pd-catalyzed pi-allyl substitution reactions. The observation that recovered butenolide ee was much less than product ee led to the development of a dynamic kinetic resolution process which was applied to the total synthesis of aflatoxin B.

### **TEACHING EXPERIENCE**

2008–present (Brandeis University): Chem 25b (Sophomore Organic lecture, 140-190 students) 135a (Advanced Organic Chemistry: Synthesis II, 8-17 students) 134b (Advanced Organic Chemistry: Synthesis I, 16 students) Chemical Biology 102a (Chemical Innovation Feeding Biological Discovery)

1998–2000 (Columbia University): *as TA*, 3 semesters organic chemistry lecture

1997–1998 (Stanford University) : *as TA*, 2 quarters organic chemistry lecture (Chem 33 and 131), 1 quarter analytical chemistry lab (Chem 134)

1998 (Stanford University): *as Head TA*, 1 quarter organic chemistry lab (Chem 130/132)

## HONORS AND AWARDS

Summer 1997: Pfizer Summer Undergraduate Research Fellowship

January 2005 – December 2007: NIH NRSA Postdoctoral Fellowship F32-AI063976

2013 Thieme Chemistry Journal Award

2013 NSF CAREER Award

2015 Strage Award for Aspiring Young Science Faculty

2015 Waltzer Award for Teaching

2017-2019 Sigma Xi Distinguished Lectureship

## PROFESSIONAL SOCIETIES

09/2000 – 09/2004, 03/2010–*present*: Member, American Chemical Society

03/2010–*present*: Member, American Society for Biochemistry and Molecular Biology

05/2019–*present*: Member, RNA Society

## PROFESSIONAL SERVICE

### *Review of Grant Proposals:*

04/2009 – ACS Petroleum Research Fund DNI proposals

04/2012 – NIH ZAI1-JBS-A M1 Special Emphasis Panel (“CHAVI-ID” HIV vaccine program)

08/2012 – ACS Petroleum Research Fund DNI proposals

2013 – NSF electronic review for Macromolecular/Supramolecular/Nanochemistry program

03/2014 – NIH/CSR *ad hoc* member, VACC study section (HIV/AIDS Vaccines)

11/2014 – NIH ZAI1 DR-A (J2) 1 Special Emphasis Panel (“Innovation for HIV Vaccine Discovery” RFA)

03/2015 – NIH ZRG1 BCMB-R (50) R Special Emphasis Panel (“Facile Methods and Technologies for Synthesis of Biomedically Relevant Carbohydrates” RFA)

06/2015 – NIH/CSR *ad hoc* member, SBICA study section (Synthetic and Biological Chemistry)

11/2015 – NIH/NIAID *ad hoc* member, AIDSRC study section (K grants for HIV research)

11/2015 – NIH/CSR *ad hoc* member, VACC study section (HIV/AIDS Vaccines)

2016 – NSF review panel for Chemical Synthesis program

04/2016 NIH ZRG1 BCMB-U (50) R Special Emphasis Panel (“Facile Methods and Technologies for Synthesis of Biomedically Relevant Carbohydrates” RFA)

2017 – NSF review panel Chemical Synthesis program

02/2017 – NIH/CSR *ad hoc* member, EBIT study section (analytical/biotechnology)

03/2017 – NIH/CSR *ad hoc* member, VACC study section (HIV/AIDS Vaccines)

07/2017 – NIH/CSR *ad hoc* member, 2017/10 ZRG1 BCMB-G (02) M (Member Conflict review panel)

10/2017 – *ad hoc* member, SBCB study section (NIH CSR)

11/2017 – NIH/NIAID *ad hoc* member, ZAI1 AL-A (J2) 1 Special Emphasis Panel (HIVRAD P01 program)

3/2018 – *ad hoc* member, ZRG1 BCMB-G(10) study section (SBIR/STTR grants) (NIH CSR)

07/2018 – NIH/NIAID *ad hoc* member, ZAI1-JRR-A (S1) 1 Special Emphasis Panel (HIVRAD P01 program)

11/2018 – NIH Special Emphasis Panel ZAI1 RB-A (J1) 1 Consortia for HIV/AIDS Vaccine Development (CHAVD)

2019 – NSF review panel for Chemical Synthesis program

02/2020 – NIH/CSR *ad hoc* member, ZRG1 BCMB-G (02) (Member Conflict review panel for synthetic chemistry and chemical biology)

02/2020 – NSF review panel for Chemical Synthesis program  
06/2020 – NIH/NIAID ZAI1-MMF-X-S2 Special Emphasis Panel (Emergency awards for COVID research)  
10/2020 – NIH/CSR *ad hoc* member, SBCA study section  
11/2020 – NIH/NIAID ZAI1 JP-A (J1) Special Emphasis Panel (Engineering Immunity to HIV-1 Through Next Generation Vaccines)  
04/2021 - NIH/NIAID ZAI1 LAR-X (M2) Special Emphasis Panel (Pan-Coronavirus Vaccines)  
07/2021 – NIH/NIAID ZAI1 JBS-A (S1) Special Emphasis Panel (HIVRAD P01 Program)  
07/2022 – NIH/NIAID ZAI1 KSW-A (S1) Special Emphasis Panel (HIVRAD P01 Program)  
12/2022 - NIH/NIAID, ZAI1 VS-A (J1) 1 Special Emphasis Panel (bNAbs Susceptibility Assay Development Program)

*Conferences Organized:*

Co-organizer, Second Boston Symposium of Encoded Library Platforms (BSELP) (August 2017)  
Co-organizer, Second Northeast Glyco-Chemistry Symposium (June 2018)  
Co-organizer, Third Northeast Glyco-Chemistry Symposium (June 2019)  
Co-organizer, Fourth Northeast Glyco-Chemistry Symposium (June 2020)  
Co-organizer, Fifth Northeast Glyco-Chemistry Symposium (June 2021)  
Co-organizer, Sixth Northeast Glyco-Chemistry Symposium (June 2022)  
Co-organizer, Seventh Northeast Glyco-Chemistry Symposium (June 2023)

*Committee Chair Service in Brandeis Chemistry:*

Manager, mass spectrometry instrumentation, 2008-present  
Undergraduate Advising Head: Fall 2015-Spring 2017  
Graduate Admissions Chair: Fall 2017-Spring 2020  
Graduate Admissions co-Chair Fall 2020-Spring 2021  
Chemistry Faculty Search Committee Chair: 2018-2019 search  
Chemistry Faculty Search Committee co-Chair: 2019-2020 search

*University-wide service at Brandeis:*

Chair, *ad hoc* committee for tenure (date withheld)  
Member, Provost Search Committee, Fall 2020-Spring 2021  
Co-Chair, Dean of A&S Search Committee, Fall 2022-Spring 2023  
Faculty Representative to the Board of Trustees, (elected for Fall 2021-Spring 2024)  
Member, Task Force on Academic Administrative Structure (Spring 2023)

Referee work for journals: *Nat. Commun.*; *Nat. Chem.*; *Nat. Chem. Biol.*; *Cell Rep.*; *Cell Chem. Biol.*; *iScience*; *ACS Central Science*; *J. Am. Chem. Soc.*; *Angew. Chem.*; *Chem. Sci.*; *Chem – Eur. J.*; *J. Org. Chem.*; *Biochemistry*; *Retrovirology*; *Org. Lett.*; *Tetrahedron Lett.*; *Bioorg. Med. Chem. Lett.*; *ACS Catalysis*; *Eur. J. Med. Chem.*, *Vaccines*, *PLoSOne*; *J. Nucleic Acids*; *Carbohydrate Research*.

## RESEARCH SUPPORT

*Active:*

NIH R21-AI174956 (4/2023-3/2025) “Mannosidase-stabilized glycan immunogens for elicitation of high mannose patch antibodies.” PI: Krauss, I. J. Award amount: \$446,875

NIH R01-AI090745 (12/2018-11/2023) “Glycopeptide evolution targeting antibodies of the PGT128 lineage.” PI: Krauss, I. J., co-I: Nemazee, D. Award amount: \$2,235,887. (2,070,551 to Krauss)

NIH R01-GM127920 (9/2018-7/2023) “Evolving New Glycosaminoglycan Mimetics.” PI: Hsieh-Wilson, L., Co-I: Krauss, I. J.) Award amount to Krauss: \$397,734

NIH R01 GM142041 (9/1/2020-8/31/2025) “Ubiquitin-independent targeted protein degradation.” PI: Lizbeth Hedstrom, Co-I Krauss, I. J.. Award amount to Krauss: \$33,030

*Completed:*

NIH R21-AI140030 (5/15/2018-5/14/2020) “Directed Evolution of Zika EDE Glycoantigens.” PI: Krauss, I. J. Award amount: \$452,974

ACS Petroleum Research Fund Doctoral New Investigator Award 51975-DNI1 (01/2012-08/2015)  
“Stereoselective Homoallylation of Aldehydes and Related Compounds” \$100,000 Total Costs

NSF CAREER CHE-1253363 (06/2013-05/2018) “Stereoselective Homoallylation and Homocrotylation”  
Award amount: \$550,000

NIH R03-AI136720 (01/2018-12/2019) “The Effect of Extended Antigen Release on Carbohydrate Specificity of the Antibody Response.” PI: Krauss, I. J. Award amount: \$162,500

NIH R01-AI113737 (07/2014-06/2019, in no-cost extension) “Design of Immunogens to Elicit PGT122-Like Antibodies” PI: Krauss, I.J., co-I: Nemazee, D. Award amount: \$2,174,374 (\$1,874,222 to Krauss)

Brandeis SPROUT Grant (Case # 2018-034) “Vaccines for Targeting HIV Sugars”; PI: Krauss, I. J. Award amount: \$20,000.

Brandeis SPROUT Grant (Case # 2019-040) “Sulfur-stabilized HIV Vaccine Antigens”; PI: Krauss, I. J. Award amount: \$30,000.

Brandeis SPROUT Grant (2020) “Mannosidase inhibitors in HIV vaccination”; PI: Krauss, I. J. Award amount: \$10,000.

## **POSTDOCTORAL AND GRADUATE PUBLICATIONS**

1. “Highly Regioselective and Diastereoselective Directed Hydroformylation of Allylic Ethers: A New Approach to Propionate Aldol Synthesis” **Krauss, I. J.**; Wang, C. C. Y.; Leighton, J. L. *J. Am. Chem. Soc.* **2001**, *123*, 11514–5.
2. “Highly Practical and Enantioselective Cu-Catalyzed Conjugate Addition of Alkylzinc Reagents to Cyclic Enones at Ambient Temperature” **Krauss, I. J.**; Leighton, J. L., *Org. Lett.* **2003**, *5*, 3201–3.
3. “Total Synthesis of Isomigrastatin” **Krauss, I. J.**; Mandal, M.; Danishefsky, S. J. *Angew. Chem. Int. Ed.* **2007**, *46*, 5576–9.
4. “Fully Synthetic Carbohydrate HIV Antigens Designed on the Logic of the 2G12 Antibody” **Krauss, I. J.**; Joyce, J. G.; Finnefrock, A. C.; Song, H. C.; Dudkin, V. Y.; Geng, X.; Warren, J. D.; Chastain, M.; Shiver, J. W.; Danishefsky, S. J. *J. Am. Chem. Soc.* **2007**, *129*, 11042–4.

5. "Total Synthesis of Spirotenuipesines A and B" Dai, M.; **Krauss, I. J.**; Danishefsky, S. J. *J. Org. Chem.* **2008**, *73*, 9576–83.
6. "An Oligosaccharide-based HIV-1 2G12 Mimotope Vaccine Induces Carbohydrate-specific Antibodies that Fail to Neutralize HIV-1 Virions" Joyce, J. G.; **Krauss, I. J.**; Song, H. C.; Opalka, D. W.; Grimm, K. M.; Nahas, D. D.; Esser, M. T.; Hrin, R.; Feng, M.; Dudkin, V.; Chastain, M. C.; Shiver, J. W.; Danishefsky, S. J. *Proc. Natl. Acad. Sci. USA*, **2008** *105*, 15684–9.
7. "A New Model for the Presentation of Tumor-Associated Antigens and the Quest for an Anticancer Vaccine: A Solution to the Synthesis Challenge via RCM" Jeon, I.; Lee, D.; **Krauss, I. J.**; Danishefsky, S. J. *J. Am. Chem. Soc.* **2009**, *131*, 14337–44.
8. "Diverted Total Synthesis Leads to the Generation of Promising Cell-Migration Inhibitors for Treatment of Tumor Metastasis: In vivo and Mechanistic Studies on the Migrastatin Core Ether Analog" Oskarsson, T.; Nagorny, P.; **Krauss, I. J.**; Perez, L.; Mandal, M.; Yang, G.; Ouerfelli, O.; Xiao, D.; Moore, M. A. S.; Massague, J.; Danishefsky, S. J. *J. Am. Chem. Soc.* **2010**, *132*, 3224–8.
9. "Confirmation of the Structures of Synthetic Derivatives of Migrastatin in the Light of Recently Disclosed Crystallographically Based Claims" Nagorny, P. **Krauss, I.**; Njardarson, J. T.; Perez, L. Gaul, C.; Yang, G.; Ouerfelli, O.; Danishefsky, S. J. *Tetrahedron Lett.* **2010**, *51*, 3873–5.

## PUBLICATIONS AT BRANDEIS

10. "Enzyme-Instructed Molecular Self Assembly Confers Nanofibers and A Supramolecular Hydrogel of Taxol" Gao, Y.; Kuang, Y.; Guo, Z.-F.; Guo, Z.; **Krauss, I. J.**; Xu, B. *J. Am. Chem. Soc.* **2009**, *131*, 13576–7.
11. "A Concise Asymmetric Synthesis of the Bromophycolide A and D Skeleton" Lin, Hongkun; Pochapsky, S. S.; **Krauss, I. J.** *Organic Lett.* **2011**, *13*, 1222–5.
12. "Multivalent Glycocluster Design Through Directed Evolution" MacPherson, I. S.; Temme, J. S.; Habeshian, S. M.; Felczak, K.; Pankiewicz, K.; Hedstrom, L.; **Krauss, I. J.** *Angew. Chem. Int. Ed.* **2011**, *50*, 11238–11242. (*Selected as a "Hot Paper"*)
13. "Homoallylboration and Homocrotylboration of Aldehydes" Pei, W.; **Krauss, I. J.** *J. Am. Chem. Soc.* **2011**, *133*, 18514–18517. (*Highlighted in Synfacts 2012*, *8*, 203).
14. "Enantioselective Homocrotylboration of Aliphatic Aldehydes" Lin, H.; Pei, W.; Wang, H.; Houk, K. N.; **Krauss, I. J.** *J. Am. Chem. Soc.* **2013**, *135*, 82–5. (*Highlighted in Synfacts 2013*, *9*, 315, and in *Organic Synthesis Highlights*, <http://www.organic-chemistry.org/Highlights/2014/24February.shtm>).
15. "Directed Evolution of 2G12-Targeted Nonamannose Glycoclusters by SELMA" Temme, J. S.; Drzyzga, M. G.; MacPherson, I. S.; **Krauss, I. J.** *Chem. - Eur. J.* **2013**, *19*, 17291-5.
16. "High Temperature SELMA: Evolution of DNA-Supported Oligomannose Clusters Which Are Tightly Recognized by HIV bnAb 2G12" Temme, J. S.; MacPherson, I. S.; Decourcey, J. F.; **Krauss, I. J.** *J. Am. Chem. Soc.* **2014**, *136*, 1726-9 (*reviewed in F1000 Prime*).
17. "Directed Evolution of Multivalent Glycopeptides Which Are Tightly Recognized by HIV Antibody 2G12" Horiya, S.; Bailey, J.; Guillen-Schlippe, Y. V.; Temme, J. S.; **Krauss, I. J.** *J. Am. Chem. Soc.*, **2014**, *136*, 5407-15 (*Highlighted in C&E News*, 3/31/2014, *reviewed in F1000 Prime*).

18. "Boron Carboxylate Catalysis of Homoallylboration" Dugas, G. J.; Lam, Y.; Houk, K. N.; **Krauss, I. J.**, *J. Org. Chem.* **2014**, *79*, 4277-84 (*Selected as Featured Article*).
19. "Recent Strategies Targeting HIV Carbohydrates in Vaccine Design" Horiya, S.; MacPherson, I. S.; **Krauss, I. J.** *Nat. Chem. Biol.* **2014**, *10*, 990-999.
20. "Glycocluster ligand selection using SELMA - SElection with Modified Aptamers" Temme, J. S.; **Krauss, I. J.** *Curr. Protoc. Chem. Biol.* **2015**, *7*, 73-92.
21. "Enantioselective *syn* and *anti* Homocrotylation of Aldehydes: Application to Formal Synthesis of Spongidepsin" Lin, H.; Tian, L.; **Krauss, I. J.** *J. Am. Chem. Soc.* **2015**, *137*, 13176-13182.
22. "Antibody Recognition of HIV and Dengue Glycoproteins" **Krauss, I. J.** *Glycobiology* **2016**, *26* (8), 813-819.
23. "Synthesis of Multivalent Glycopeptide Conjugates that Mimic an HIV Epitope" Bailey, J.; Nguyen, D. N.; Horiya, S.; **Krauss, I. J.** *Tetrahedron*, **2016**, *72* (40), 6091-6098.
24. "DNA Display of Folded RNA Libraries: Removing the Reverse Transcription Step from RNA Selection" MacPherson, I. S.; Temme, J. S.; **Krauss, I. J.** *Chem. Comm.* **2017**, *53*, 2878-2881.
25. "Structural Characterization of Early Michaelis Complexes in the Reaction Catalyzed by (+)-Limonene Synthase from *Citrus sinensis* using Fluorinated Substrate Analogs" Kumar, R. P.; Morehouse, B. R.; Matos, J. O.; Malik, K.; Lin, H.; **Krauss, I. J.**; Oprian, D. D. *Biochemistry* **2017**, *56*, 1716-1725.
26. "Directed Evolution of Glycopeptides Using mRNA Display" Horiya, S.; Bailey, J. K.; **Krauss, I. J.** *Meth. Enzymol.* **2017**, *597*, 83-141.
27. "Stereoselective Homocrotylation of Aldehydes: Enantioselective Synthesis of Allylic-Substituted Z/E-Alkenes." Tian, L.; **Krauss I. J.** *Org. Lett.* **2018** *20*, 6730-6735.
28. "Oligomannose glycopeptide conjugates elicit antibodies targeting the glycan core rather than its extremities" Nguyen, D. N.; Xu, B.; Stanfield, R. L.; Bailey, J. K.; Horiya, S.; Temme, S. J.; Leon, D. R.; LaBranche, C. C.; Montefiori, D. C.; Costello, C. E.; Wilson, I. A.; **Krauss, I. J.** *ACS Cent. Sci.* **2019**, *5*, 237-249.
29. "Direct evidence for an enzyme generated LPP intermediate in (+)-limonene synthase using a fluorinated GPP substrate analog." Morehouse, B. R.; Kumar, R. P.; Matos, J. O.; Yu, Q.; Bannister, A.; Malik, K.; Temme, J. S.; **Krauss, I. J.**; Oprian, D. D. *ACS Chem. Biol.* **2019**, *14*, 2035-2043.
30. "The Impact of Sustained Immunization Regimens on the Antibody Response to Oligomannose Glycans." Nguyen, D. N.; Redman, R. L.; Horiya, S.; Bailey, J. K.; Xu, B.; Stanfield, R. L.; Temme, S. J.; LaBranche, C. C.; Wang, S.; Rodal, A. A.; Montefiori, D. C.; Wilson, I. A.; **Krauss, I. J.** *ACS Chem. Biol.* **2020**, *15*, 789-798.
31. "Mechanism Underlying Anti-Markovnikov Addition in the Reaction of Pentalene Synthase." Matos, J. O.; Kumar, R. P.; Ma, A. C.; Paterson, M.; **Krauss, I. J.**; Oprian, D. D. *Biochemistry* **2020**, *59*, 3271-3283.

32. "Synthesis of Mannosidase-Stable Man<sub>3</sub> and Man<sub>4</sub> Glycans Containing S-linked Man $\alpha$ 1 $\rightarrow$ 2Man Termini." Neralkar, M.; Tian, L.; Redman, R. L.; **Krauss, I. J.** *Org. Lett.* **2021**, *23*, 3053-3057.
33. "Directed Evolution of 2'-Fluoro-Modified, RNA-Supported Carbohydrate Clusters that Bind Tightly to HIV Antibody 2G12" Redman, R. L.; **Krauss, I. J.** *J. Am. Chem. Soc.* **2021**, *143*, 8565-8571.
34. "An Optimized Synthesis of Fmoc-L-homopropargyl Glycine" Polyak, D.; **Krauss, I. J.** *J. Org. Chem.* **2022**, *87*, 3841-3844.
35. "Homoallylboration of Aldehydes: Stereoselective Synthesis of Allylic-Substituted Alkenes and E-Alkenes" Polyak, D.; Xu, B.; **Krauss, I. J.** *Org. Lett.* **2022**, *24*, 4656-4659
36. "A Mechanistic Switch from Homoallylation to Cyclopropylcarbinylation of Aldehydes" Polyak, D.; Xu, B.; Lee, W.; Houk, K. N.; **Krauss, I. J.** *Org. Lett.* **2022**, *24*, 4660-4664
37. "Large-Scale Synthesis of Man<sub>9</sub>GlcNAc<sub>2</sub> High-Mannose Glycan and the Effect of the Glycan Core on Multivalent Recognition by HIV Antibody 2G12" Neralkar, M.; Xu, B.; Horiya, S.; **Krauss, I. J.** *ACS Infect. Dis.* **2022**, *8*, 2232-2241.
38. "In vitro selection of cyclized, glycosylated peptide antigens that tightly bind HIV glycan patch antibodies" Bailey, J. K.; Neralkar, M.; Horiya, S.; Krauss, I. J." *manuscript in preparation*
39. "Profiling of Methionine Amino Peptidase (MAP) Substrate Specificity Using mRNA Display Selection" Bailey, J. K.; Turra, R. J.; Neralkar, M.; Horiya, S.; Krauss, I. J." *manuscript in preparation*

### **LECTURES (invited, unless otherwise noted)**

1. Stonehill College (October 2008)
2. University of Southern Maine (November 2008)
3. Merrimack College (October 2010)
4. Carbohydrates Gordon Research Conference (June 2011)
5. Colgate University (November 2011)
6. Dartmouth College (January 2012)
7. ACS Symposium on Host-Pathogen Interactions, Carbohydrate Section (March 2012, 243<sup>rd</sup> ACS meeting)
8. Brown University (April 2012)
9. Hobart William and Smith College (April 2012)
10. ACS Organic Division "Young Academic Investigators Symposium" (August 2012, 244<sup>th</sup> ACS meeting)
11. Tufts University (October 2012)
12. University of Massachusetts, Dartmouth (November 2012)
13. Rhode Island College (November 2012)
14. University of Rhode Island (February 2012)
15. Hokkaido University, Sapporo, Japan (February 2012)
16. Bioorganic Chemistry Gordon Research Conference (June 2013)
17. Carbohydrates Gordon Research Conference (June 2013)
18. Organic Reactions and Processes Gordon Research Conference (July 2013)
19. Ra Pharma (July 2013)
20. Emmanuel College Merck Lecture (September 2013)
21. Clark University (October 2013)
22. Merrimack College (October 2013)
23. MRSEC Seminar, Brandeis University (November 2013)
24. Northeastern University (Dec 2013)



25. University of Pittsburgh (February 2014)
26. University of Pennsylvania (February 2014)
27. Temple University (February 2014)
28. University of Delaware (February 2014)
29. Texas A&M University (February 2014)
30. Bridgewater State University (February 2014)
31. Young Investigators in Glycoscience Symposium (March 2014, 247<sup>th</sup> ACS meeting)
32. Columbia University (April 2014)
33. University of North Carolina, Chapel Hill (April 2014)
34. University of California, Irvine (April 2014)
35. University of Texas, Austin (April 2014)
36. University of Colorado, Boulder (April 2014)
37. Colorado State University (April 2014)
38. University of California, Davis (April 2014)
39. University of California, Santa Barbara (April 2014)
40. University of California, Berkeley (April 2014)
41. Memorial Sloan-Kettering Cancer Center (May 2014)
42. The Scripps Research Institute, La Jolla (May 2014)
43. Bowdoin College (May 2014)
44. Brandeis University, Chemistry Department (September 2014)
45. St. Anselm College (October 2014)
46. University of Georgia / Complex Carbohydrate Research Center (October 2014)
47. University of Michigan, Midwest Carbohydrate Symposium (October 2014)
48. PerkinElmer (scheduled for October 2014)
49. Middlebury College (November 2014)
50. University of Michigan, Department of Medicinal Chemistry, (February 2015)
51. Biochemistry/Biophysics Friday Seminar, Brandeis University (March 2015)
52. "Frontiers in Glycoscience" Symposium (March 2015, 249<sup>th</sup> ACS Meeting, Denver, CO)
53. "Carbohydrate Synthesis for Medicinal Chemistry and Biology" Symposium (August 2015, 250<sup>th</sup> ACS Meeting, Boston, MA)
54. SUNY Albany (October 2015)
55. "Carbohydrate Recognition in Health and Disease" Symposium #342 (Pacifichem, December 2015)
56. Albert Einstein College of Medicine, Department of Biochemistry (April 2016)
57. University of New England, Biddeford, ME (September 2016)
58. Glaxo Smith-Kline, Waltham, MA (September 2016)
59. Merck Discovery Chemistry, Kenilworth, NJ (October 2016)
60. Rhode Island College (October 2016)
61. Purdue University (November 2016)
62. Symposium on Recent Technological Advances of Nucleic acid Ligands Against Extracellular Targets, Pittconn, 2017 (March 2017)
63. David Gin Award Symposium (April 2017, 253<sup>rd</sup> ACS Meeting, San Francisco, CA)
64. Second Boston Symposium of Encoded Library Platforms (August 2017)
65. Worcester State (September 2017)
66. University of Arizona (January 2018)
67. Northern Michigan University (March 2018)
68. Bridgewater State University (March 2018)
69. Bates College (April 2018)
70. Midwest Carbohydrate and Glycobiology Symposium at Michigan State (September 2018)
71. Merrimack College (October 2018)
72. Wheaton College (October 2018)
73. Georgia Southern University (November 2018)

74. Boston Glycobiology Discussion Group, Harvard University (January 2019)
75. Lamar University (March 2019)
76. Chemical Glycobiology of Glycopeptides Symposium (March 31<sup>st</sup>, 257<sup>th</sup> ACS Meeting, Orlando, FL)
77. Wayne State University (planned March 2020, canceled due to Coronavirus)
78. Rhode Island College (planned April 2020, canceled due to Coronavirus)
78. Peking University (planned April 2020, canceled due to Coronavirus)
79. Nankai University (planned April 2020, canceled due to Coronavirus)
80. Huazhong USTC (planned April 2020, canceled due to Coronavirus)
81. Shanghai Jiao Tong University (planned April 2020, canceled due to Coronavirus)
82. Great Lakes Translational Glycomics Symposium (planned for August 2020, canceled due to Coronavirus)
83. St. Joseph's University, virtual visit (October 2020)
84. Rhode Island College, virtual visit (November 2020)
85. "Oligosaccharide Synthesis, Conjugation, and Vaccine Antigenicity" Symposium (August 2022 National ACS Meeting, Chicago, IL)
85. New Reactions and Methodology Section (self-submitted) (August 2022 National ACS Meeting, Chicago, IL)
86. University of Maryland, College Park, (scheduled for May 2023)

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