#### 1. Coupling Transcription and DNA Repair with a dsDNA-Tracking Motor

2. Post-Translational Modification of Tubulin by Tubulin Tyrosine Ligase (TTL)



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Lodish et al., Molecular Cell Biology

MILLER SPREAD COURTESY OF DR. SARAH FRENCH

### A Stalled RNA Polymerase Can Cause a Molecular Pile-up





AFM of transcription complexes on irradiated template

Trautinger et al., (2005) *Molecular Cell* 19, 247-254 Adapted from: <u>http://www.cawlocal584.com/humour.html</u>

# 3000 RNA Polymerase molecules/cell 10-20 DNA Polymerase III molecules/cell



Lodish et al., Molecular Cell Biology

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# **RNAP Stalling Leads to Genomic Instability**

- RNAP/DNAP collisions result in double-strand breaks
- DNA repair shapes the mutational landscape of cancer cells
- DNA repair is the **main** mechanism underlying the development of cancer in all living organisms
- Also essential for the mechanism of chemotherapeutic agents (irofulven)
- Essential for the etiology of accelerated-aging diseases (Cockayne Syndrome)

Deaconescu *et al.,* (2012) *Trends Biochem, Sci.37(12)* 543-552 Nudler, E. (2012) *Cell* 149(7) 1438-1445 Nik-Zainal *et al.,* (2012) *Cell* 149(5) 994-1007



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# Mfd (mutation frequency decline)

#### **Radiation-Induced Mutations and Their Repair**

Evelyn M. Witkin

Science, New Series, Vol. 152, No. 3727 (Jun. 3, 1966), 1345-1353.

- mutant is UV sensitive
- 3-fold lower rate of excision of pyrimidine dimers



# Mfd is the TRCF (transcription-repair coupling factor)

Proc. Natl. Acad. Sci. USA Vol. 88, pp. 11574–11578, December 1991 Biochemistry

# *Escherichia coli mfd* mutant deficient in "mutation frequency decline" lacks strand-specific repair: *In vitro* complementation with purified coupling factor

(transcription-repair coupling/UV mutagenesis/SOS response/nonsense suppressors)

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Contributed by Evelyn M. Witkin, September 26, 1991

Selby et al., (1991) PNAS (88) 11574-11578

# Main Players



(?) (?)  $(\circ)$   $(\circ)$ 

# **TRCF Mediates Transcription-Coupled Repair (TCR)**



Question

# How are the different TRCF activities integrated?

## **Multiple Activities Are Integrated in a Multi-Domain Protein**





Park et al. (2002) Cell 109, 757

# The D2-D7 Clamp Is Inhibitory to Translocation on Naked DNA



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# **UvrA Recruitment Requires Opening of the D2/D7 Clamp**



# **SAXS Probes Structure in Solution**



Adapted from Petoukhov, M., EMBL lecture

Measure isotropic intensity distribution and radially average I(q) ullet<u>;</u>; ?? <u>;</u>; • Extract shape parameters:  $R_q$  (low q data) ?? **4**€ ≥ 4€ D<sub>max</sub> (maximum intramolecular distance) ?? ??? **S3bS** 

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Deaconescu et al., PNAS (2012) 109 (9): 3353-3358

#### ??????????pau????g???h??tig?u?????



 Reconstruct 3D-envelope using *ab initio* algorithms starting with gas phase of "dummy atoms/residues" in a spherical search volume followed by minimization and fit against the scattering curve

#### SYSTEM IS UNDERDETERMINED, MULTIPLE SOLUTIONS ARE POSSIBLE

# The D2/D7 Clamp is Maintained in Solution in the Nucleotide-Free State



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## The Catalytic Cycle Reorganizes Interdomain Contacts



Deaconescu et al., PNAS (2012) 109 (9): 3353-3358

# The Catalytic Cycle Reorganizes Interdomain Contacts



Deaconescu et al., PNAS (2012) 109 (9): 3353-3358

#### **ATP•TRCF** Simulations are Robust and Reproducible

 Use the PDB of apo-TRCF as starting model (input) to seed the simulation process

Ab initio simulation PDB-seeded simulation



# Probing Clamp Opening Using Interdomain Disulfide Engineering



TRCF Variant	Crosslinking w/ ATP	Crosslinking w/ UvrA
TRCF-D7:RID (open, more extended)	+++	+++
TRCF-D2:D7 (closed, more compact)		

# The D2/D7 Clamp Is Ideally Positioned to Restrain the TRCF Motor and Provide Spatiotemporal Control



Prevents UvrA Binding Compromises DNA binding Impairs ATP turnover (in the absence of RNAP) Still releases RNAP off templates due to stimulation by the RNAP elongation complex  $\rightarrow$ 

# UvrA may be recruited after release of RNAP/transcript from the elongation complex

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# Post-translational modification of tubulin by a tubulin tyrosine ligase (TTL)

(a collaboration with the Roll-Mecak Lab, NIH)



http://schulmanlab.jhu.edu/research.html

http://micro.magnet.fsu.edu/cells/microtubules/ microtubules.html

#### **Microtubules are Dynamic**



**Conde & Caceres (2009) Nat Rev Neuroscience** 

Nature Reviews Neuroscience

### **Microtubules are Heterogeneous**



Carsten Janke & Jeannette Chloë Bulinski

Nature Reviews Molecular Cell Biology (2010) 12, 773-786

# Tubulin Tyrosine Ligase (TTL) Modifies the α-Tubulin Tail



Garnham & Roll-Mecak. Cytoskeleton (2012) Volume 69, Issue 7, pages 442-463

#### **A Putative Mechanism for Tyrosination**



# TTL-How Does It Work, and Why Is It Important?

- Important for organization of neural networks
- Microtubules in dendrites are enriched in Tyr tubulin
- TTL suppression leads to cancer
- TTL prefers to tyrosinate free tubulin heterodimer over tubulin that has been incorporated into a lattice (e.g. microtubules)
- TTL inhibits tubulin polymerization



# **TTL Binds Tubulin Not only Through Its Tail**



X. tropicalis TTL (Monomer)

Szyk et al., (2012) NSMB

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K_d (tubulin) ~ 1 \muM
K_d (peptide) ~ 144 \muM
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# What is the basis for this substrate specificity?



(lateral, between protofilaments)

#### Hypothesis TTL binds to a surface that is buried in the polymeric tubulin form

### The TTL-Tubulin Complex is Elongated



# The TTL Contacts Primarily α-Tubulin



#### Is the tubulin heterodimer flipped?

- AUC confirmed complex formation and 1:1 stoichiometry
- From frictional ratio → aspect ratio is 3.8 (versus 2.2 for tubulin) denoting an elongated structure

Szyk et al., (2012) NSMB

### Antibodies against α-Tubulin Inhibit Tyrosination



# TTL May Modulate the Partitioning of Heterdimeric versus Polymeric Tubulin and Tune Motors and End-Binding Proteins



Szyk et al., (2012) NSMB

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Dr. Aga Szyk

HHMI HOWARD HUGHES MEDICAL INSTITUTE

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