

Force as a variable to study biological reactions

Biochemistry under stress!

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QB Lecture-1

Lecture outline

- Lecture-1:
 - Why force??
 - Mechanical processes in biology
 - Motor proteins
 - Mechanosensing
 - Experimental techniques
- Lecture-2:
 - Force → free energy landscape
 - What we can learn from using force to study a mechanical process: RNA unfolding
- Lecture-3:
 - Motor enzymes and mechanochemistry
 - Mechanistic information from force-velocity studies

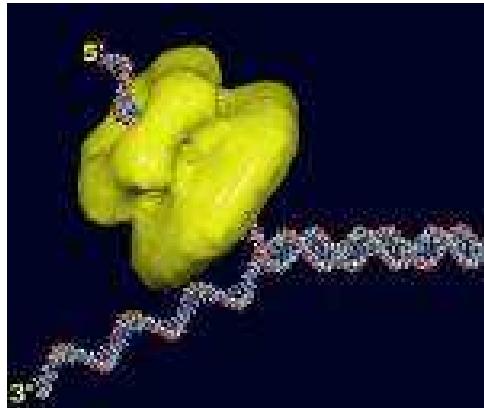
Why force??

Forces can be an **input** or an **output**

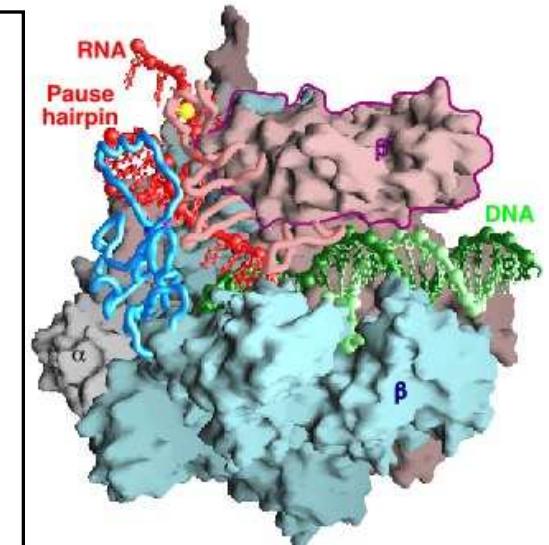
Motor proteins: force generating nanomachines
chemical (or electrochemical) energy →
mechanical work

Mechanosensing:
mechanical force → cellular signal

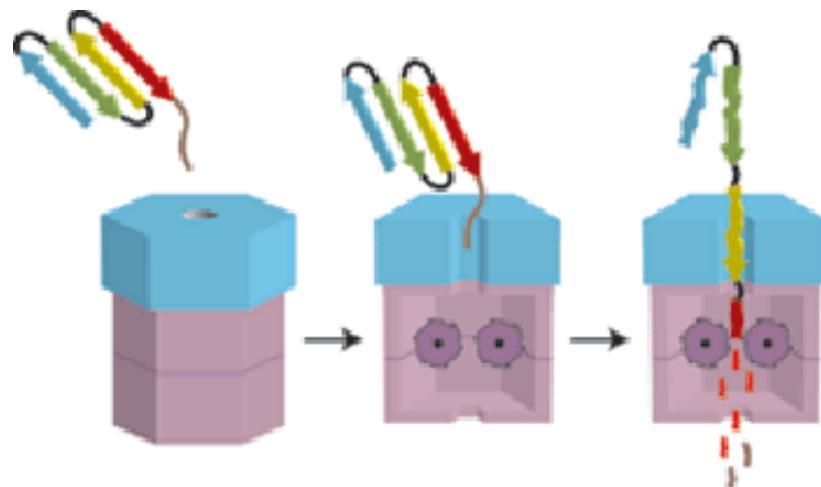
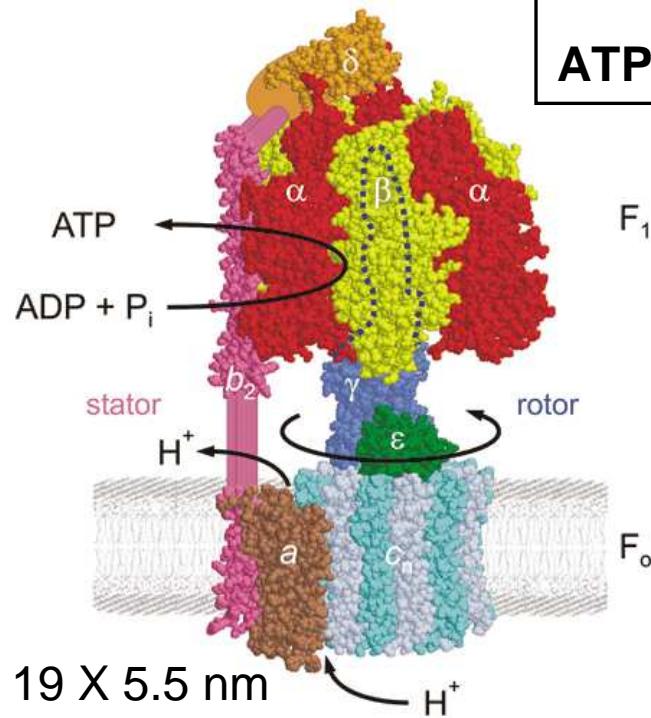
View of cell as a collection of nanomachines



Replication:	Helicase DNA polymerase
Transcription:	RNA Polymerase
Translation:	Ribosome
Degradation:	Proteosome
ATP synthesis:	F0F1 ATP synthase

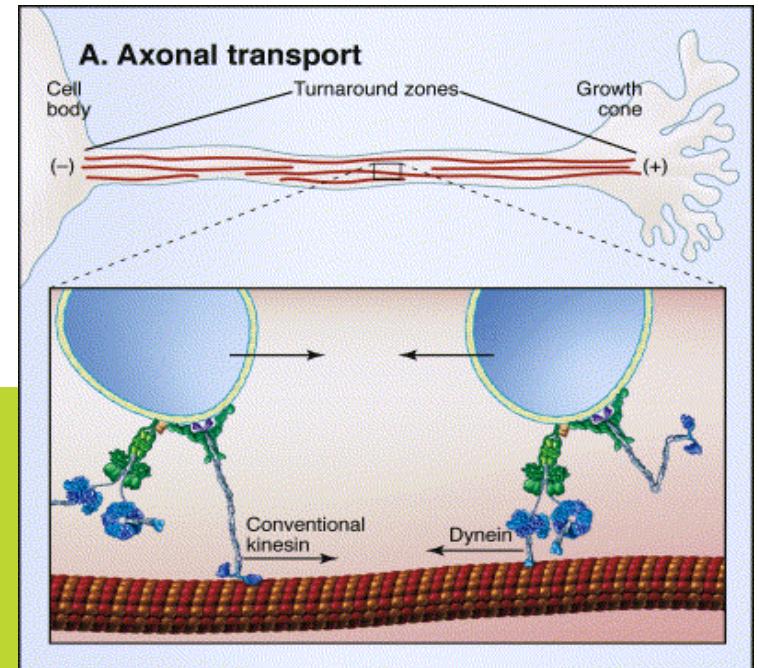
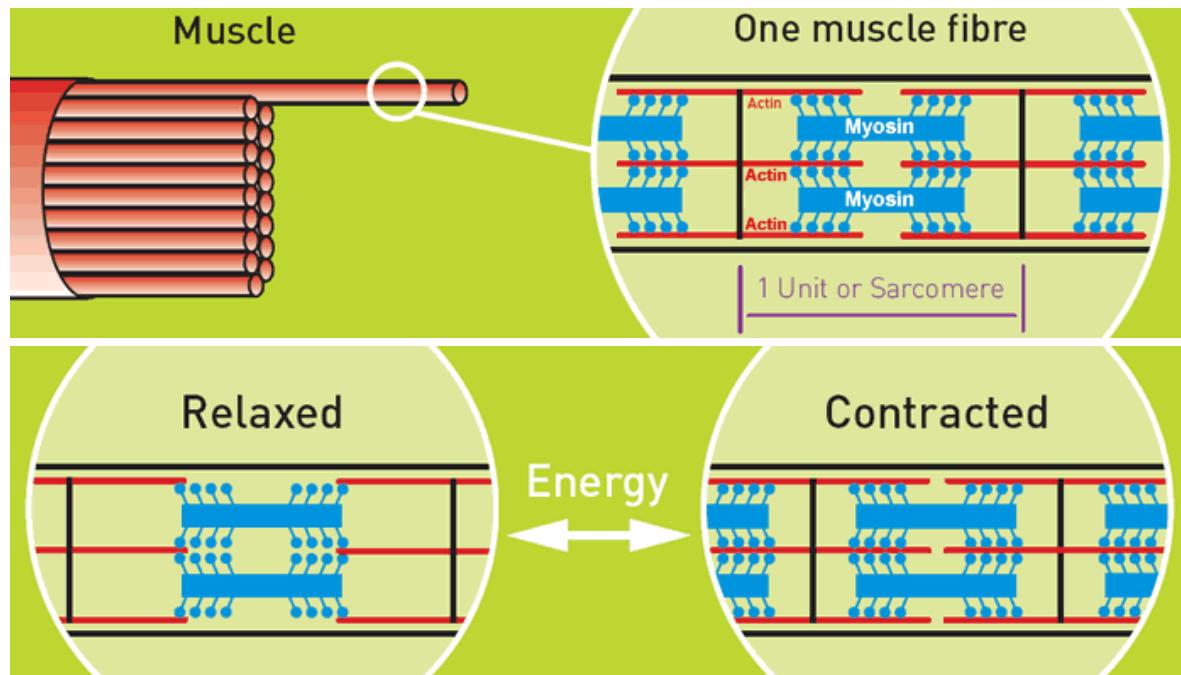


8.5 x 10.5 x 14 nm
25 pN



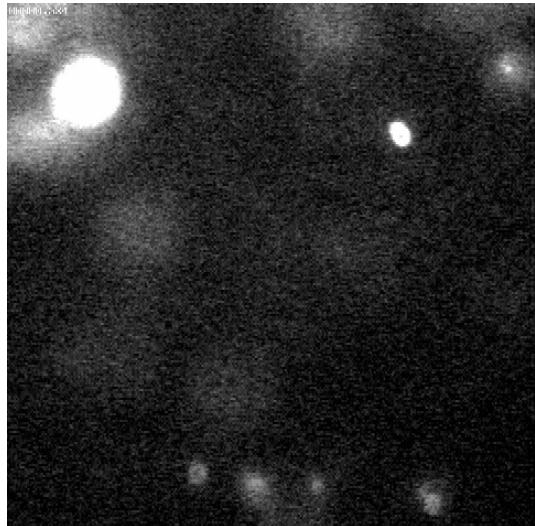
Cytoskeletal motors kinesin, dynein, myosin

- Sub-cellular motility
- Cell division
- Muscle contraction

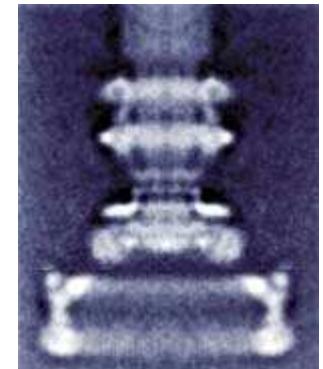
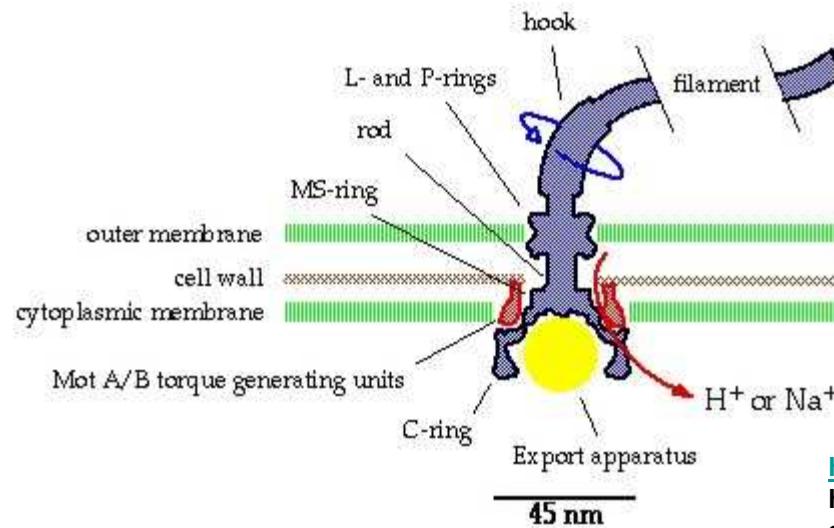


Vale, R.D. The molecular motor toolbox for intracellular transport. *Cell* 112, 467-80 (2003).

Flagella



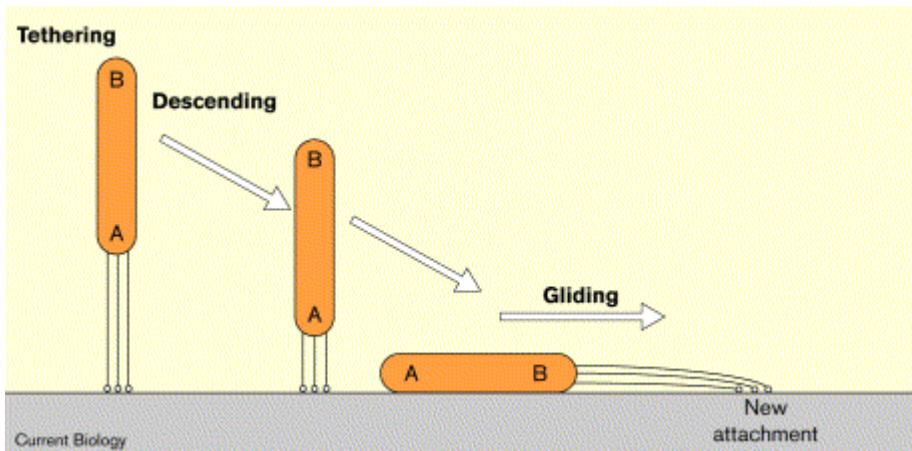
Cellular motility



<http://www.aip.org/pt/jan00/berg.htm>
<http://www.physics.ox.ac.uk/biophysics/research/flagellar.html>

Movies from <http://www.rowland.org/labs/bacteria/>

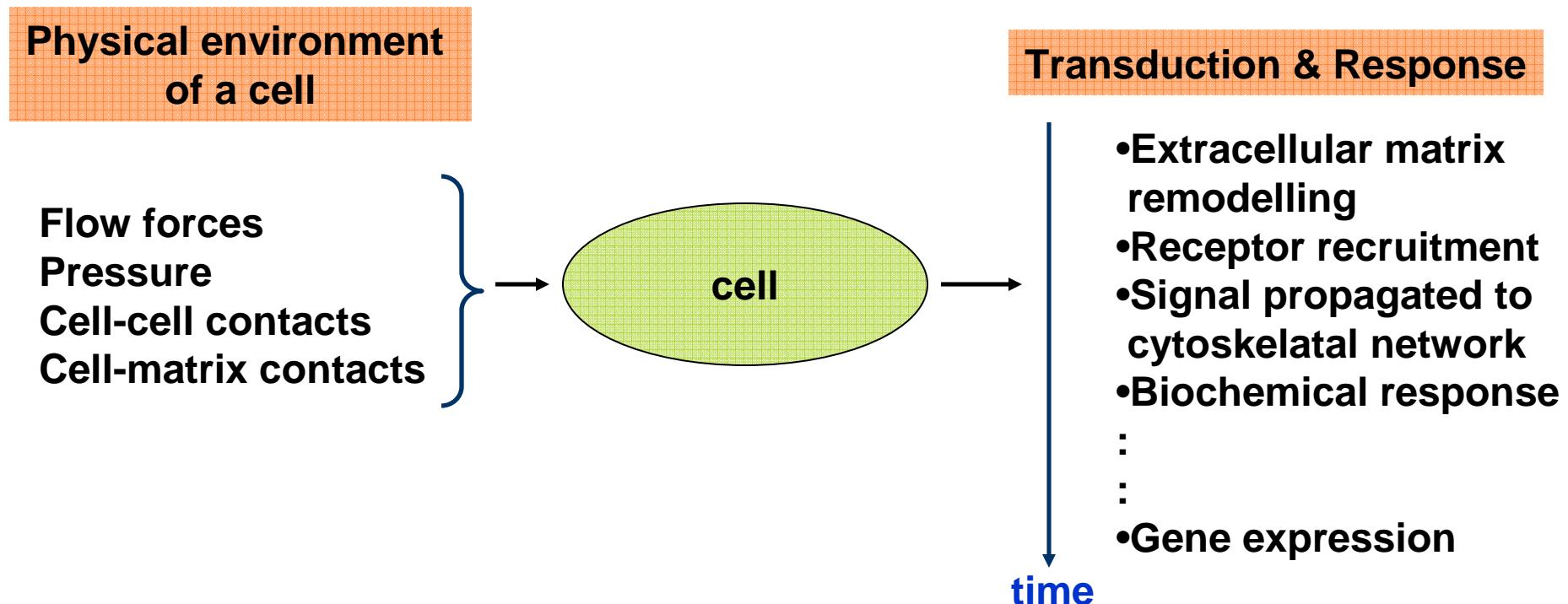
Pilus



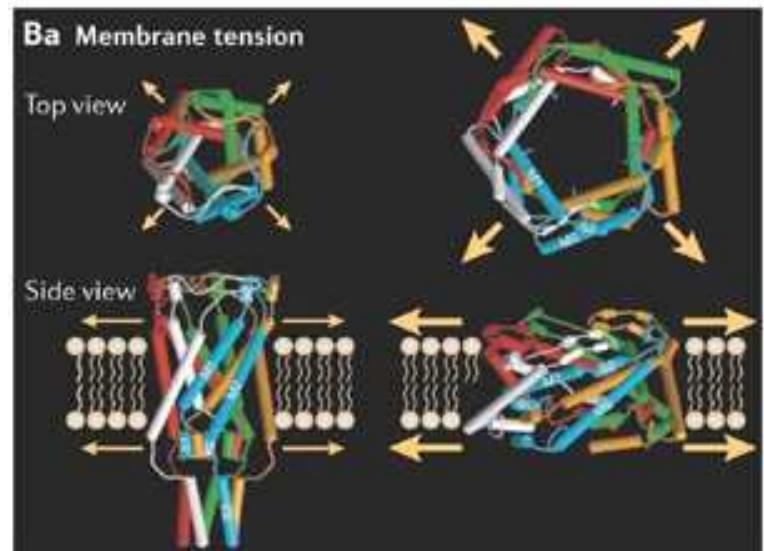
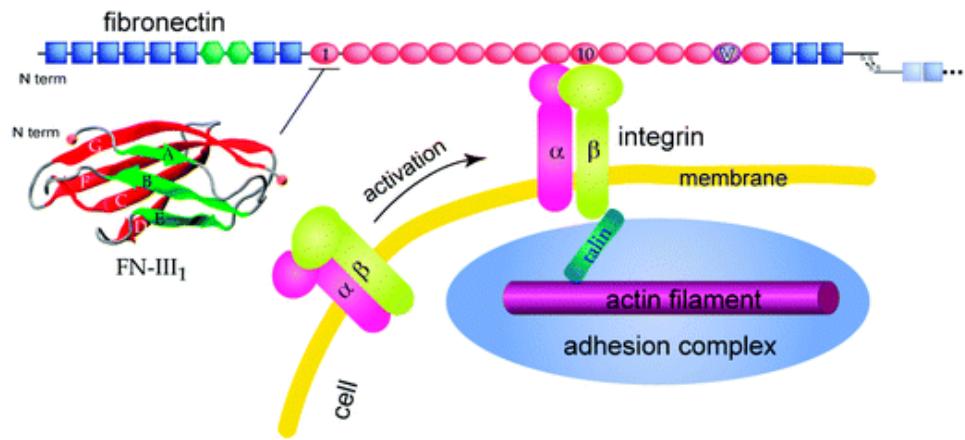
Kaiser, D. Bacterial motility: how do pili pull? *Curr Biol* 10, R777-80 (2000).

Mechanosensing, transduction and response

physical signal → primary sensing → transduction → response

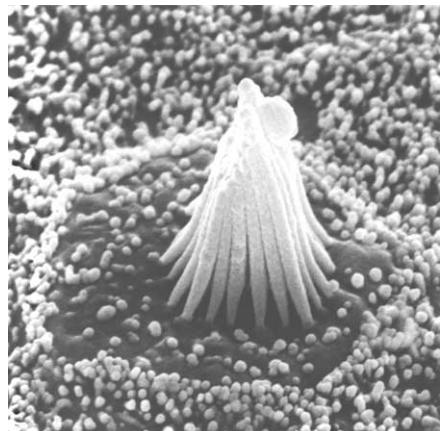
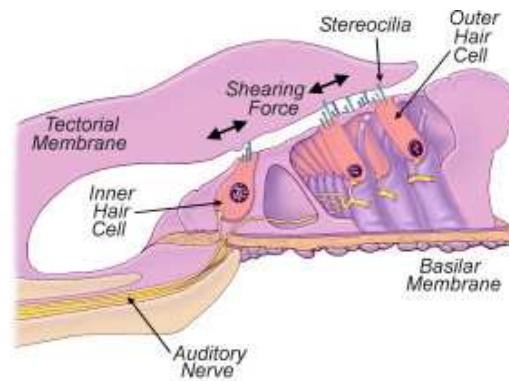
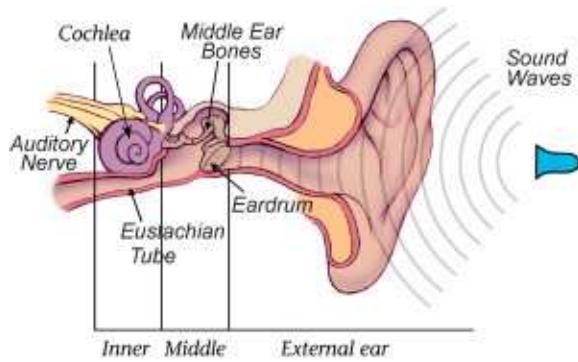


Primary sensing by “force receptors”

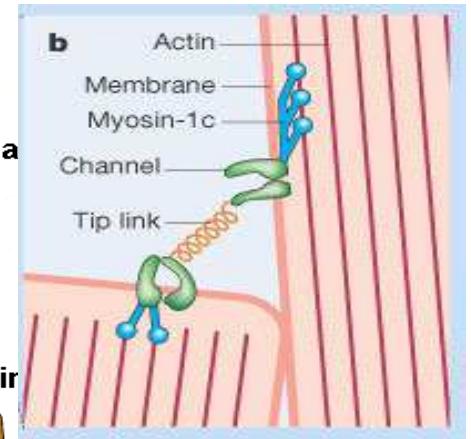
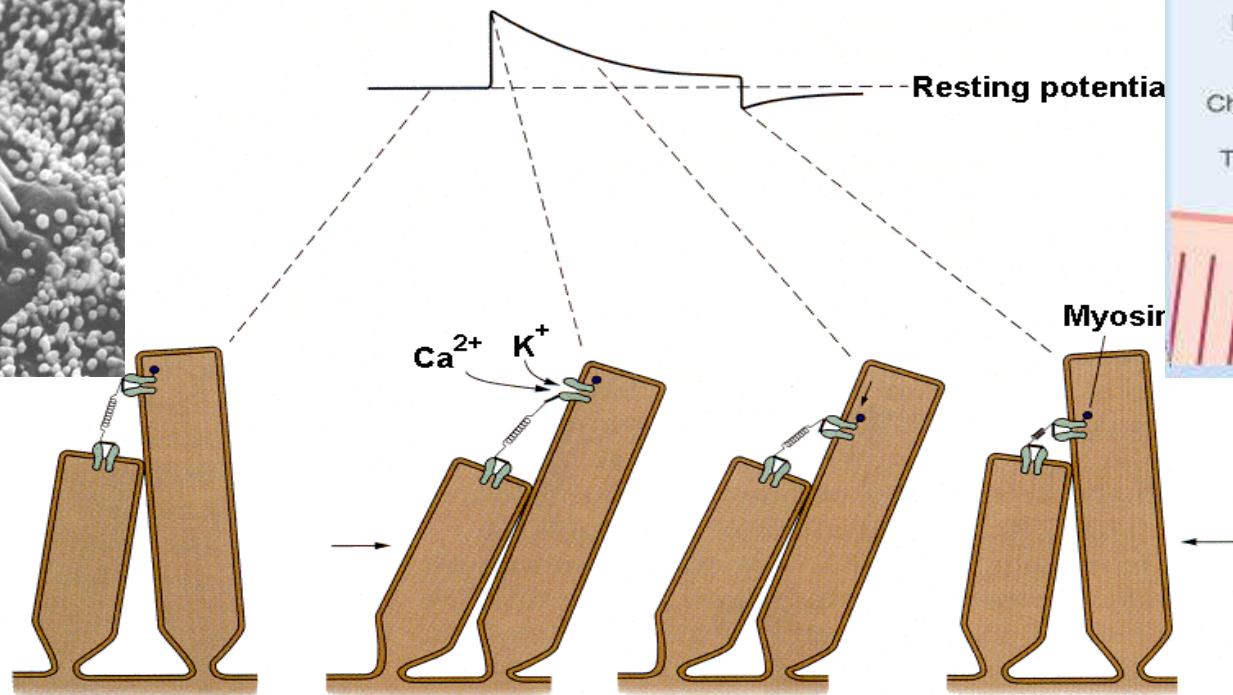


Gao, M., Sotomayor, M., Villa, E., Lee, E.H. & Schulten, K. Molecular mechanisms of cellular mechanics. *Phys Chem Chem Phys* 8, 3692-706 (2006).
Vogel, V. & Sheetz, M. Local force and geometry sensing regulate cell functions. *Nat Rev Mol Cell Biol* 7, 265-75 (2006).

Mechanosensing in hair cells

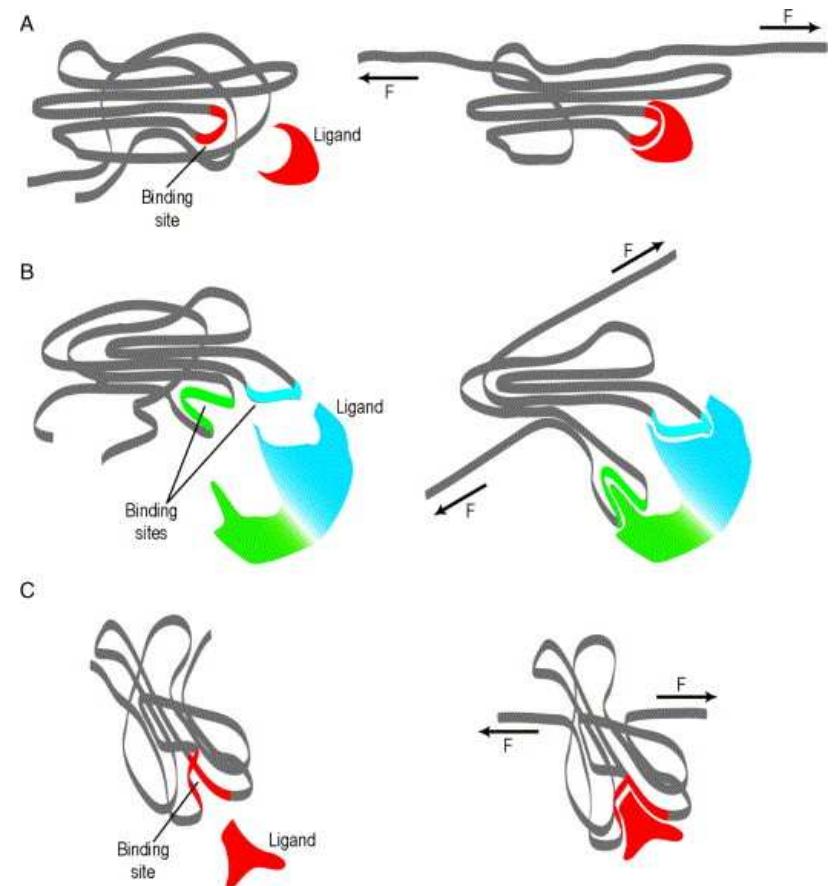
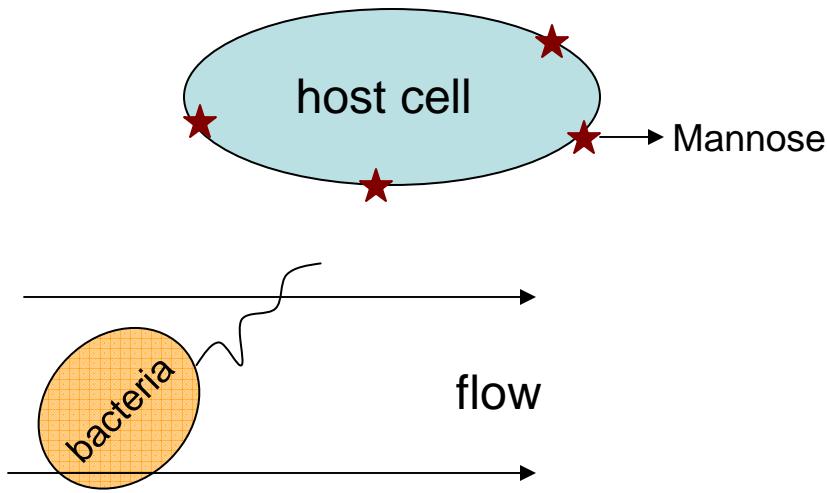


stereocilia

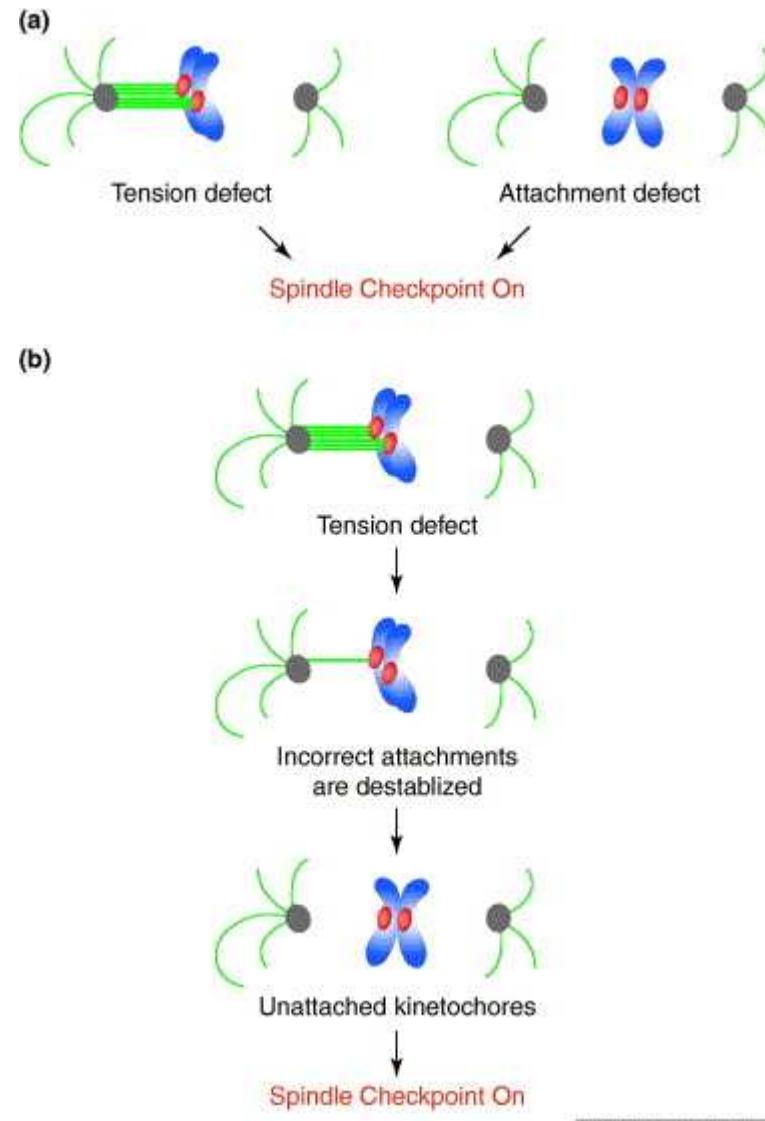
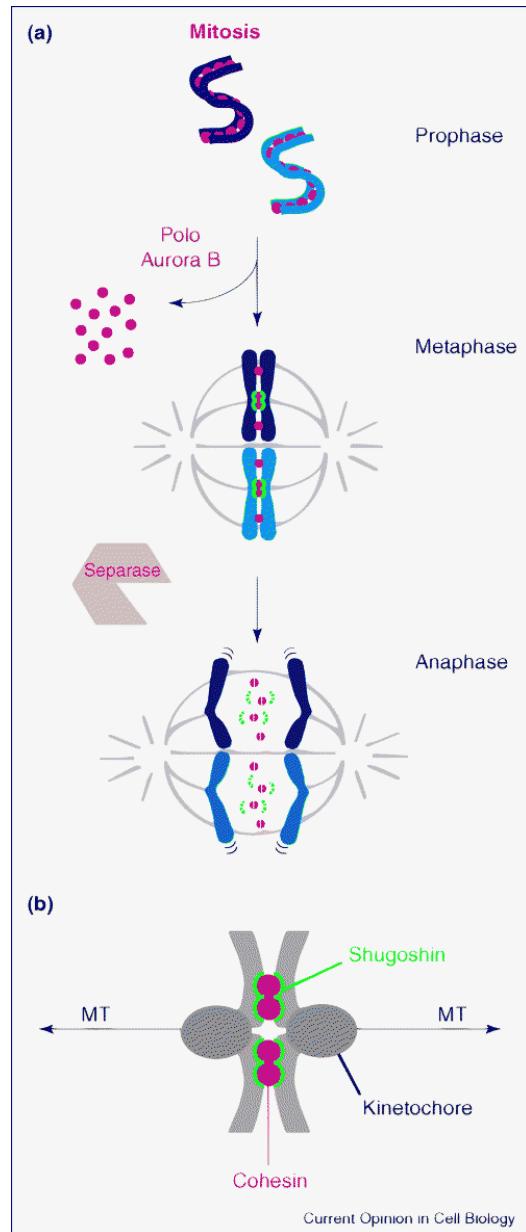


Mechanosensing by “Catch Bonds”

short → long lived attachments at increasing forces



Tension sensing during chromosome segregation

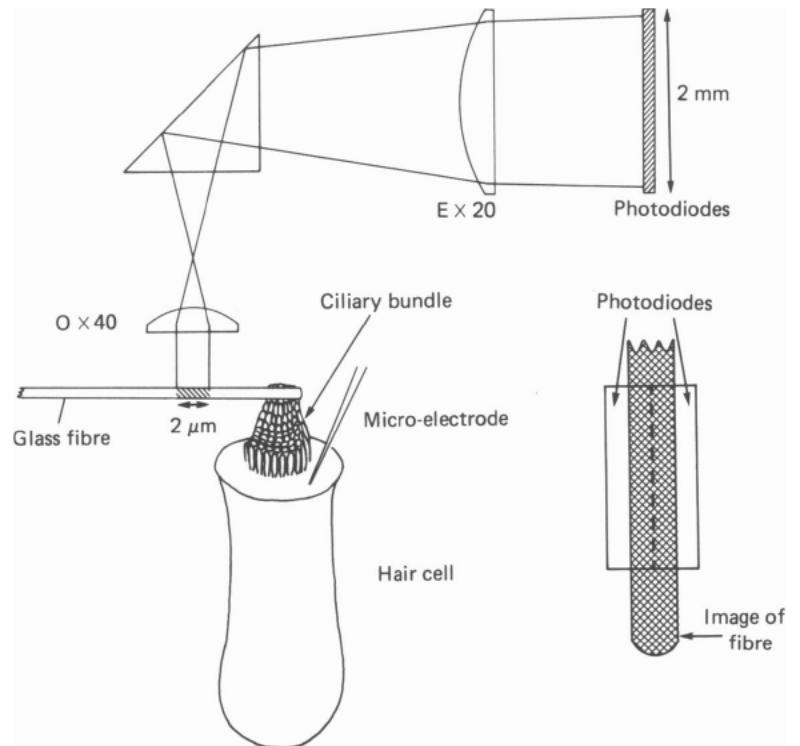
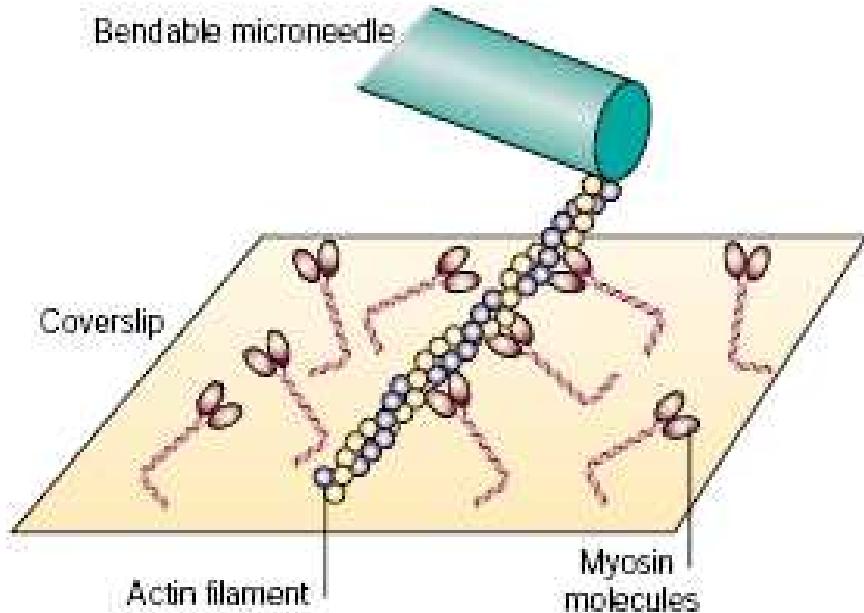


Pinsky, B.A. & Biggins, S. The spindle checkpoint: tension versus attachment. *Trends Cell Biol* **15**, 486-93 (2005).

Watanabe, Y. Shugoshin: guardian spirit at the centromere. *Curr Opin Cell Biol* **17**, 590-5 (2005).

Techniques

Microneedle or glass fibres

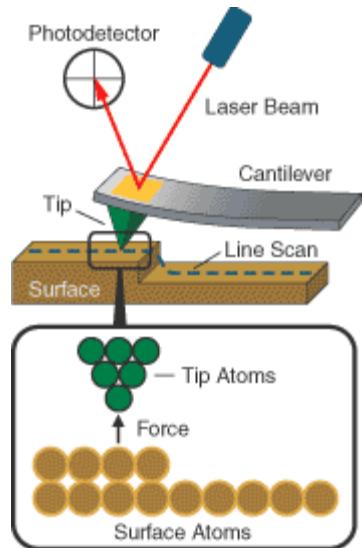
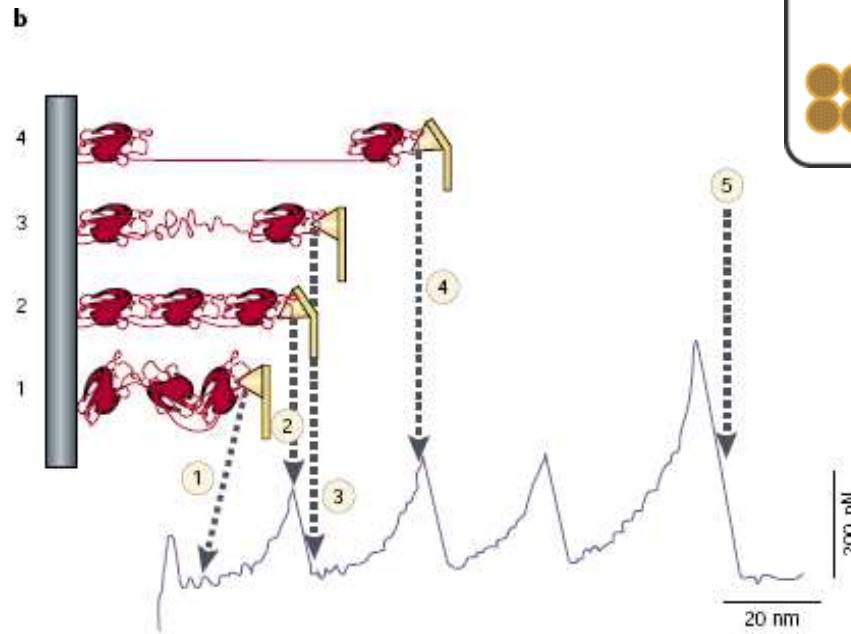
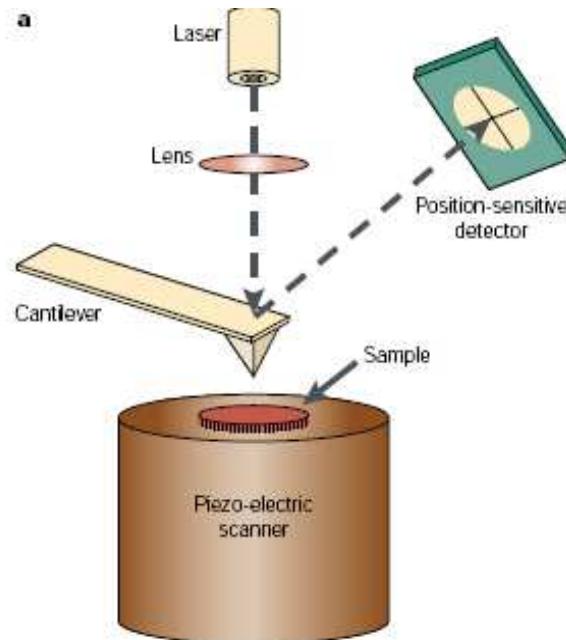


Force range: 1-100 pN

Position resolution: 1 nm

Bustamante, C., Macosko, J.C. & Wu, G.J. Grabbing the cat by the tail: manipulating molecules one by one. *Nat Rev Mol Cell Biol* 1, 130-6 (2000).
Crawford, A.C. & Fettiplace, R. The mechanical properties of ciliary bundles of turtle cochlear hair cells. *J Physiol* 364, 359-79 (1985).

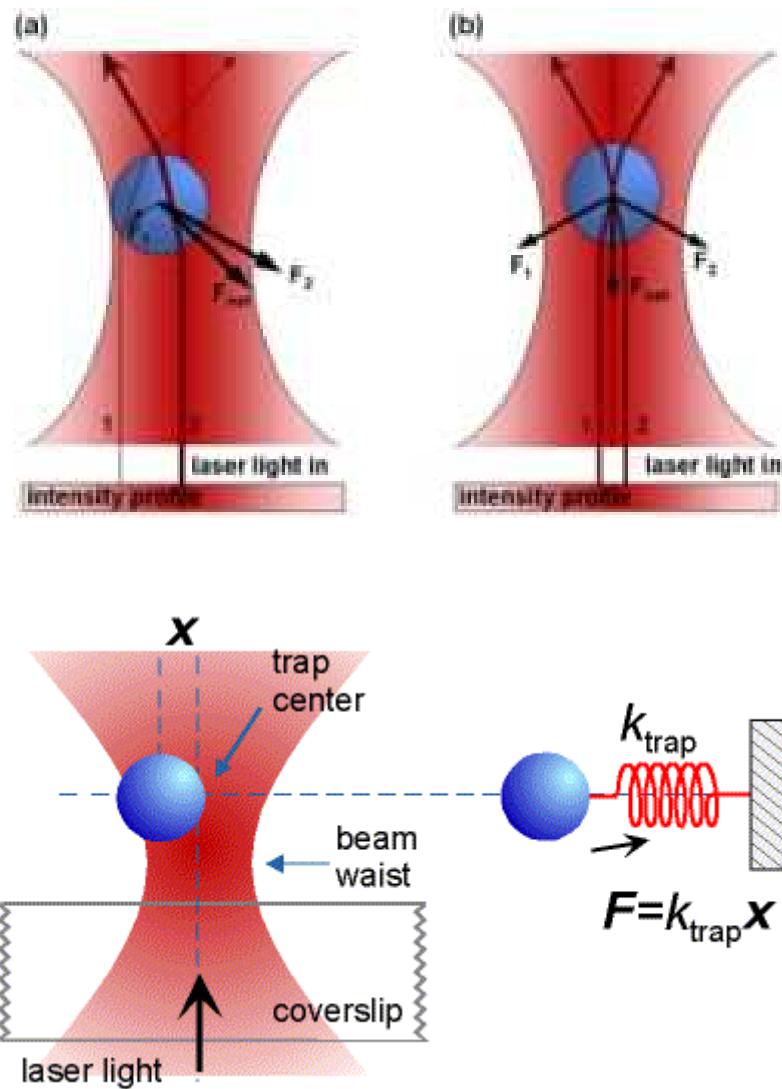
Atomic force microscopy



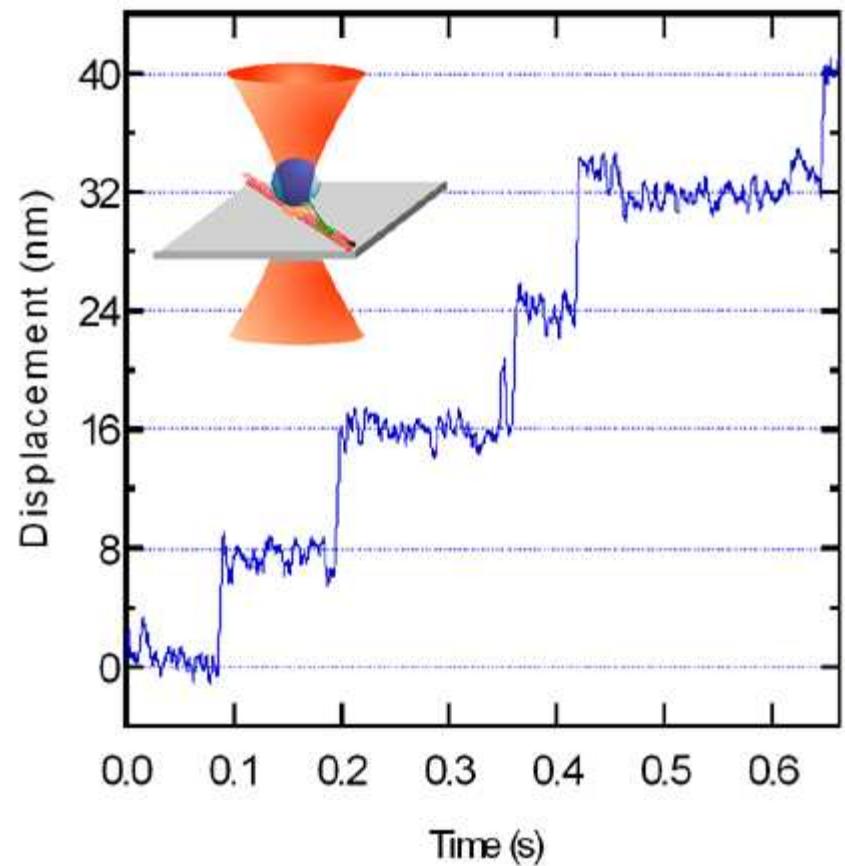
Force range: 10 pN -10 uN
Position resolution: 0.1 nm

Bustamante, C., Macosko, J.C. & Wuite, G.J. Grabbing the cat by the tail: manipulating molecules one by one. *Nat Rev Mol Cell Biol* 1, 130-6 (2000).
http://www.molec.com/what_is_afm.html

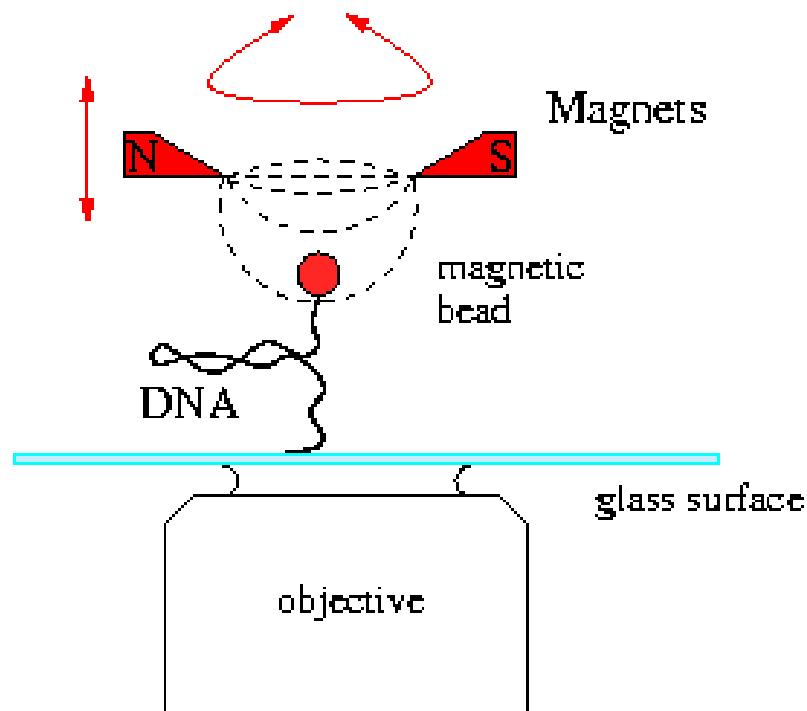
Optical tweezers



Force range: 0.1-100 pN
Position resolution: 1 nm



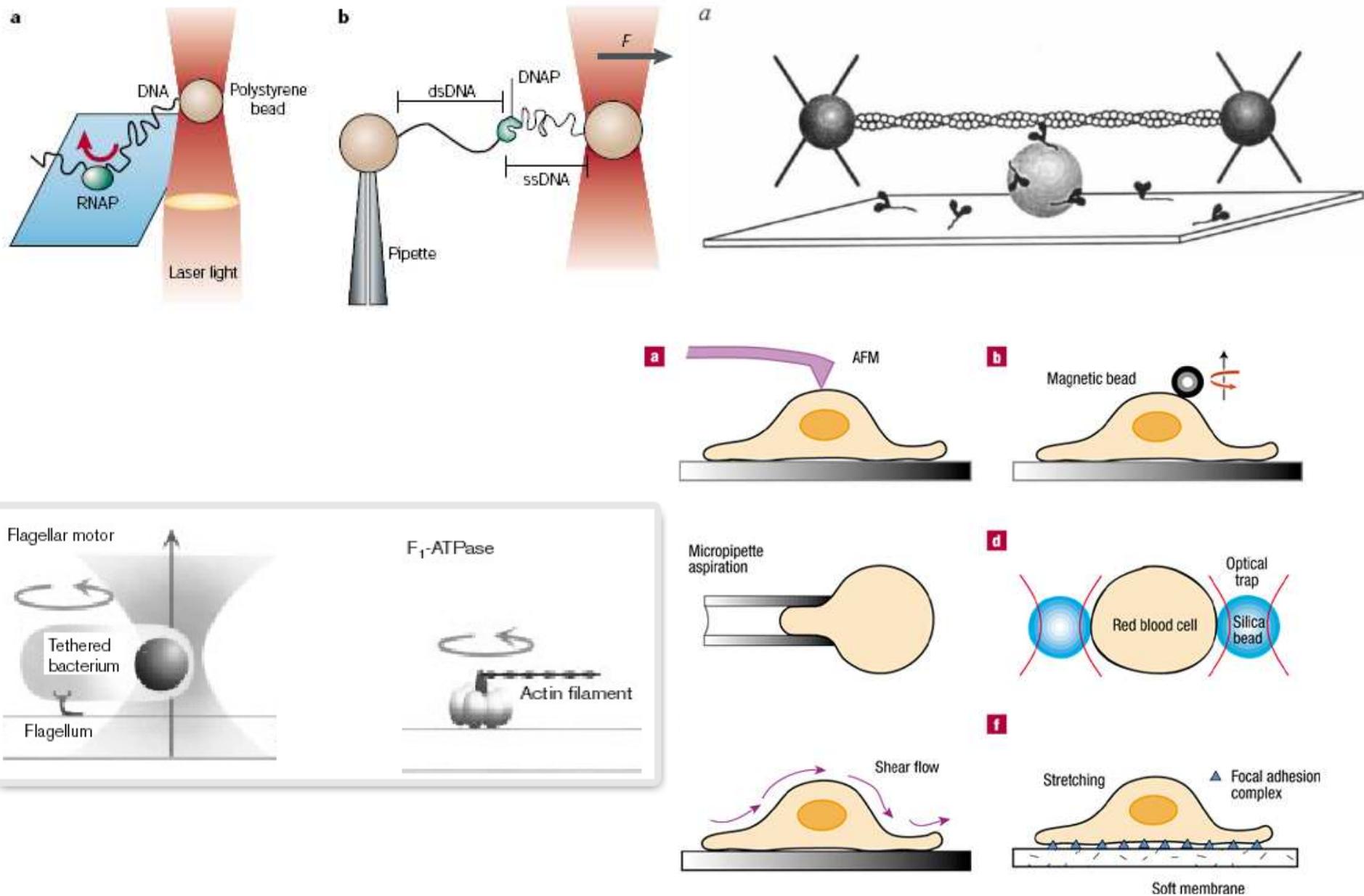
Magnetic tweezers



Force range: 0.01-10 pN
Position resolution: 10 nm

Figure from <http://www.picotwist.com/index.php?content=smb&option=mag>

Various experimental designs



A “cool” experiment !

